

A new approach for water purification from microbial pollution

E. Moosavi, V. Martirosyan, S. Ayrapetyan

Abstract—The fact that low concentrations of CO₂ have an activation effect on functional activity of microbes allows us to suggest that CO₂ could elevate the toxic effect of H₂O₂ on cells. To check this hypothesis the dependency of the toxic effect of H₂O₂ on wild type of *Escherichia coli* K-12 on soluble concentration of CO₂ in culture media was studied. The obtained data show that culture media enriched with CO₂ leads to the increase of toxic effect of H₂O₂ on microbes at both cases when pH is constant and when it changes.

Keywords— water purification, antibacterial properties, CO₂, H₂O₂, *Escherichia coli* K-12

I. INTRODUCTION

As the hazardous effects of environmental pollution on organisms are realized mostly through the water medium, the problem of water purification is considered by the UNESCO and World Health Organization (WHO) as one of the global problems of the Modern Environmental Science [7, 24]. The World Health Organization estimates that about 4.000 children die from water-born diseases every day, while United Nation has shown that more than one billion people are still without access to clean drinking water, and 2.6 billion people have no access to proper sanitation [7]. The investigation of these problems calls forth a great number of researches to be conducted to identify robust new methods of purifying water at lower prices and with less energy [2, 9]. Our research is also directed to find alternative water purification method and namely methods and compositions for disinfecting water in artificial water systems (swimming pools etc.) using CO₂/H₂O₂ combination system in different environmental factors.

Currently H₂O₂ and CO₂ are being used as tools for water purification but separately. If the molecular mechanism of “killing” effect of H₂O₂ on bacteria is well documented [5], the mechanism of “killing” effect of CO₂ on bacteria still remains discussable [6]. Although it is already shown that SCCO₂ and H₂O₂ combination make inactivate spores to high degree [10], the authors of the mentioned article do not know the mechanism

by which this inactivation takes place and also this combination has not been used as a water purification tool.

In our previous works we showed that the treatment of aqua nutrient medium by extremely low frequency electromagnetic field (ELF EMF) and infrasound (IS) could significantly modulate the growth and the development of microbes in water [21, 25]. Such treatment of water leads to the changes of CO₂ solubility in water and at the presence of oxygen it promotes the formation of reactive oxygen species (ROS) such as: singlet oxygen (¹O₂), superoxide radical (O₂⁻), peroxide anion (O₂⁻²), hydroxyl radical (OH·) and hydrogen peroxide (H₂O₂), through which the biological effects of this treatment are realized [16, 22]. On the other hand in our previous work, performed on cells of eukaryotes, it was shown that membrane proteins, determining cell membrane functional activity, are functionally in active and inactive (reserve) states. The ratio of these active and inactive molecules could be changed by the modulation of cell hydration [18, 20]. It was also shown, that metabolic poisons cause cell hydration [19] bringing to the increase of the number of functionally active receptors in the membrane [18]. On the one hand CO₂, being a strong metabolic poison, which leads to the increase of cell hydration, could serve as a potential factor able to elevate membrane sensitivity to H₂O₂ and on the other hand comparatively low concentrations of CO₂ have an activation effect on functional activity of microbes [6]. Thus, it was suggested that CO₂ could elevate the toxic effect of H₂O₂ on cell.

Therefore, working hypothesis for the present work was suggested: the activation of microbe metabolism induced by comparatively low concentrations of CO₂ could be used for the elevation of the toxic effect of H₂O₂ on microbes. This fact and data obtained from our published patent [17] let us suggest that a certain combination of both of “killer” molecules would give us opportunity to develop a novel and safe method for water purification. To check this hypothesis the following experiments were performed: *the study of different-exposure-time-dependent effects of CO₂ on growth of E. coli K-12; the study of dose-dependent effects of H₂O₂ on growth of E. coli K-12; the study of the antibacterial properties of CO₂/H₂O₂ combination on growth of E. coli K-12.*

II. PROBLEM FORMULATION

Due to increases of human population and rapid industrialization nowadays water pollution is becoming a serious problem. So, one of the global challenges of the 21st century is to find the best method for cleanup of aquatic ecosystems from microbes, which will be cheap, safe for human health and environmentally friendly. Our research

E. Moosavi is with the Department of Microbiology; Babol University of Medical Sciences, Babol, IRAN (e-mail: emosavi4000@yahoo.com)

V. Martirosyan is with the Department of Biotechnology; UNESCO Chair-Life Sciences International Postgraduate Educational Center; 31 Acharyan str., 0040 Yerevan; ARMENIA (e-mail: varsikmartirosyan@gmail.com).

S. Ayrapetyan is with the Department of Biotechnology; UNESCO Chair-Life Sciences International Postgraduate Educational Center; 31 Acharyan str., 0040 Yerevan; ARMENIA (corresponding author to provide phone: 374 10 624170; fax: 374 10 624170; e-mail: info@biophys.am).

relates to a new approach of cleaning water from microbial pollution, which will give an opportunity to solve some of the above mentioned problems. To achieve our goals we have used following methods and materials.

A. Experimental setup and apparatus

The processing apparatus used in this work contained 50 liter CO₂ vacuum tank (Fig. 1 a), the airline tubing with regulator valve (Fig. 1 b) and the CellStar CO₂ incubator (Fig. 1 c) (USA, SWJ500TV BA.), where the samples were located for CO₂ exposure. All the samples, which were incubated in CellStar incubator, were treated by CO₂ for different exposure time period. The appropriated sham tube (control tube) was also placed in the same CO₂ incubator. Compressed carbon dioxide was supplied from a gas cylinder, which was linked to the pressure vessel by a valve (Fig. 1 d). A precision manometer (Fig. 1 e) was installed in the vessel to measure the pressure. The pressure in the vacuum tank could be adjusted between 5 kPa to atmospheric pressure, and the pressure at the end of the experiment could be controlled.

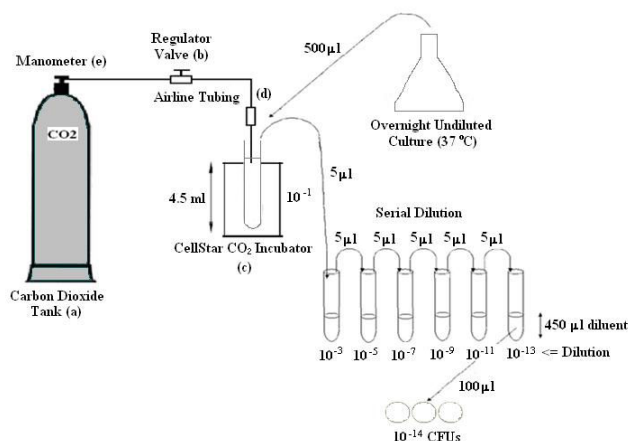


Fig. 1 A schematic blow-up of carbon dioxide into the bacterial culture and schematic representation of the serial dilution and culture.

B. Design and process of experiments

As for the experimental design, three factors were studied, which are the following: CO₂ concentration (mol l⁻¹) or exposure time (min) of CO₂, pH and H₂O₂ concentration (mmol l⁻¹). The main responses of the above mentioned factors bring to the inactivation of bacteria and the acidity of the medium. The experimental tubes containing bacterial culture were placed in the CellStar incubator at the beginning of each experiment, and the temperature was kept constant in this incubator. Then carbon dioxide (Air Liquide, 99.7% pure) was injected into the incubator at the selected pressure (atmospheric pressure). Microbial cells were treated with CO₂ for various exposure times. The apparatus allows us to inject carbon dioxide very quickly into the bacterial culture (within 10 s). The tubes in the incubator were closed and removed, and the survived cells were immediately counted (CFUs count). The inside surfaces of the incubator were wiped clean

with cotton moistened with a 70% alcohol solution before each test.

C. Bacterial strains and culture media used

We used the wild type of *Escherichia coli* K-12, which was obtained from the Armenia Collection of Microorganisms (Institute of Microbiology of the Center of Microbiology and Microbial Depository of Armenian NAS). It is a good model of organism for this work, because *E. coli* are widely distributed in waters including natural waters, particularly in tropical regions of the globe and it is closely related to other pathogenic bacteria in the enteric family (*Shigella*, *Salmonella*, etc.). During the experiment as a culture media we have used nutrient enriched broth (NEB) and nutrient enriched agar (NEA), containing: NEB + 1.5% agar (Difco), final pH 7.1 ± 0.2 at 37°C.

D. Enumeration of living microorganisms

The viability of *E. coli* K-12 was determined by counting the number of colony forming units (CFUs) for per milliliter. Measuring techniques involve indirect viable cell counts, also called plate counts, which is performed by plating out (spreading) a sample of a culture on a nutrient agar surface by triplicate plating. Each treated sample (5 µl) was diluted with 450 µl of an aqueous NEB solution before plating. The contents of the tubes were then shaken well by magnetic stirring for two min. By shaking the tubes the microorganisms dispersed in the solution. The dispersion obtained was then diluted repeatedly, and 100 µl of the appropriate dilution was plated on triplicate petri dishes containing a suitable growth medium. After plated on a suitable medium and after incubation in thermostat (37°C) each viable unit grew and formed colony and the number of CFUs was related to the viable number of bacteria in the sample. Microbial cells in the control samples were counted by the same procedures described for treated samples. The acidity of each treated sample was expressed by measuring the pH of the NEB media.

E. The method of CO₂ concentration measurement

There are several ways of determining the level of CO₂ in liquid media. Common analyze which was done for the measurement of the concentration of CO₂ comprised a reaction of a sample during which a certain amount of alkaline fluid was added, e.g. NaOH [23], to neutralise the carbon dioxide. Through titration the remaining amount of OH⁻ we can determine the amount of dissolved CO₂. So the content of CO₂ in bacterial culture media was determined by acid-base titration method (quantitative chemical analysis). The time for one titration was about 10 min. The end-point was determined by potentiometrical use of pH meter.

F. Statistical analysis

Statistical analysis was conducted using JMP software (Version 5.1.2, SAS Institute Inc., Cary, NC, USA, 2004). All

experiments were repeated at least three times. The differences among the means of treatments were tested by using *t* test.

III. PROBLEM SOLUTION

In present work we have investigated the effect of CO₂ and subsequent H₂O₂ treatment on the inactivation processes of the bacterial culture of *E. coli* K-12. The increase of CO₂ concentration in liquid and the subsequent addition of H₂O₂ made the inactivation processes of bacterial culture more effective. To check out the above mentioned hypothesis the following studies were done.

A. The study of different-exposure-time-dependent effects of CO₂ on growth of *E. coli* K-12

Carbon dioxide has a dual physiological role in microorganisms since it can both stimulate and inhibit cell development [3]. At comparatively low concentrations of CO₂ the latter has strong stimulatory effect on the growth and development of microbes [6] while in higher concentrations and in the case of longtime exposure it has inhibitory effect on the growth and development of microbes [14]. Various hypotheses have been proposed to explain the microbicidal activity of carbon dioxide. CO₂ dissolves in water to form carbonic acid. Thus, dissolved CO₂ acts by lowering the pH of the medium, and the resulting acidity leads to a disturbance of some biological systems within cells. It was therefore suggested that microbial inhibition was due to an alteration in the properties of cell (membrane, cytoplasm, enzymes, etc.) [12]. However, a reduction in the pH of the medium is not sufficient to account for the antimicrobial action of CO₂, since it shows a specific inhibitory effect, which is greater than that of the other acids used to lower medium acidity (hydrochloric acid, phosphoric acid, etc.). These acids do not penetrate the microbial cells as easily as carbon dioxide [8]. Therefore, it was suggested that the comparatively low concentration of CO₂-induced activation of the metabolism of microbes could serve as a convenient method for the increase of the sensitivity of microbes to the toxic effect of low concentration of H₂O₂.

To find out direct and indirect effects of CO₂ on growth of *E. coli* K-12 we have studied the effects of CO₂ in constant and changing pH conditions. Since CO₂ dissolves in aqueous solutions, to form an acid we need to keep constant the level of pH in order to avoid the effects of latter on bacterial killing processes. We have kept the level of pH constant by adding NaOH (0.1 mol l⁻¹) during CO₂ treatment. It should be mentioned that we have kept the value of pH constant (pH=6), because in case of this value the growth of *E. coli* bacterial culture is maximal [11]. So to find out the direct effect of CO₂ we have studied the influence of time-dependent exposure of CO₂ on the growth of *E. coli* K-12 at constant pH (pH=6) (CO₂=0.55 mol l⁻¹) (Fig. 2). The same figure also illustrates the indirect effect of CO₂ as this kind of treatment was done in different pH values (Fig. 2).

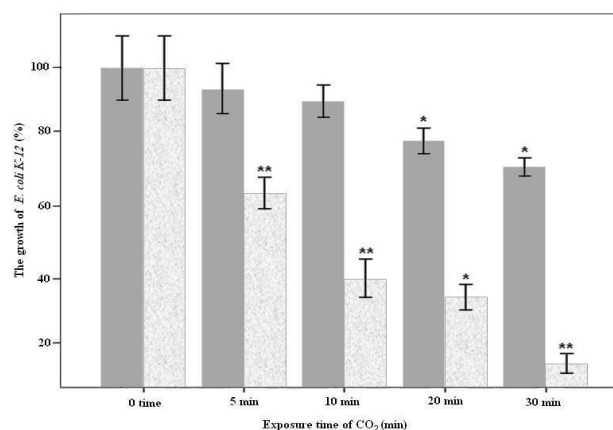


Fig. 2 The ratio of killed *E. coli* K-12 (%) depending on exposure time of CO₂ (0, 5, 10, 20 and 30 min) compared with control sample (0 time), columns illustrate the direct (■) and indirect (▨) effects of CO₂ on the growth of bacterial culture at constant (pH=6; CO₂=0.55 mol l⁻¹) and at different pH conditions, correspondingly: 0 time (pH=7.29; CO₂=0.11 mol l⁻¹), 5 min (pH=6.34; CO₂=0.51 mol l⁻¹), 10 min (pH=5.66; CO₂=0.81 mol l⁻¹), 20 min (pH=5.65; CO₂=0.84 mol l⁻¹), 30 min (pH=5.61, CO₂=0.85 mol l⁻¹). The p value of the treatments were 0.01(**), 0.05(*) and the number of experiment was 3.

We have checked the rate of inactivation of the *E. coli* K-12 as a function depending on exposure time. The ratio of surviving cells after treatment with CO₂ was compared with the ratio of cells in the control culture (0 time) and the rate of inactivation was expressed in percent (Fig. 2). After CO₂ exposure samples were incubated for 2 h at 20°C, then plating was done and the samples were incubated at 37°C thermostat for 22 h. The number of living cells was determined by measuring the number of colonies grown on each agar-medium. Zero time of treatment represents the control samples or zero time of the treatment means that the sample was not exposed to CO₂. The number of cells in the suspension was counted in nutrient agar plates with the surface plating method (CFUs for per ml of sample 10¹⁰). The inactivation of microbial cells was enhanced by the increase of time of exposure.

As it can be seen the indirect effect of CO₂ at different pH conditions on microbes is higher than the direct effect of CO₂ when pH is constant (pH=6, CO₂=0.55 mol l⁻¹) (Fig. 2), but the direct effect of CO₂ on microbes has also significant decreasing effect as compared with control group (0 time). These data correspond with the literature data that the toxic effect of CO₂ on microbes could not be explained only by acidification medium, probably because of its higher membrane permeability has direct poisoning effect on cell metabolism. Thus, the obtained data show that CO₂-induced depression of microbe growth was more pronounced than in case of equivalent decrease of pH by adding HCl acid. This conclusion completely corresponds with the previous literature data on this subject [4, 8]. The different drop in pH, compared with the control sample (pH=7.29), was observed in all of the experiments (Fig. 2). Since CO₂ dissolves in aqueous solution

to form an acid pH was lowered after each treatment, but with further increases in experimental condition values, the measured pHs was not changed significantly, which means that the end-point of acidity was not a function of the exposure time. So we can see that the initial inhibition is increased with the elevation of exposure time of CO₂ (Fig. 2).

B. The study of dose-dependent effects of H₂O₂ on the growth of *E. coli* K-12

It is known that the response of *E. coli* to H₂O₂ highly depends on its concentration. Low or high concentrations of the oxidant bring to the production of different species leading to cell death via two different mechanisms: at low concentrations microbes are forming filaments, while at higher concentrations microbes are shrinking [1, 5, 13]. In the present work we have checked the possibility to elevate the toxic effect of comparatively low doses of H₂O₂ on microbes by preliminary increase of the metabolic activity of microbes by pre-incubation of the latter in the aqua medium enriched with CO₂. Before all these procedures we studied the dose-dependent effects of H₂O₂ on the growth of *E. coli* K-12 (Fig. 3).

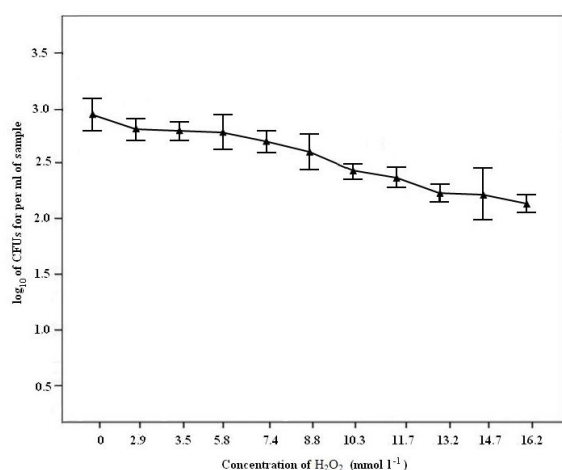


Fig. 3 The dose-dependent effects of H₂O₂ on the growth of *E. coli* K-12. The values are the average of three experiments with the standard deviation.

For hydrogen peroxide challenge, a fresh hydrogen peroxide solution was prepared immediately before starting the experiments and added to the bacterial culture bringing to the corresponding concentration of hydrogen peroxide in aqua media. Samples were incubated for 2 h at 20°C. The number of cells in the suspension with H₂O₂ concentration of several dilutions from 0 to 16.2 mmol l⁻¹ (or 0.055-0.01%) was counted in nutrient agar plates with the surface plating method, plates were inoculated at 37°C for 22 h and colonies were counted to determine the number of survived cells after H₂O₂ exposure: then the average and standard deviation of each point were calculated. After obtaining the average value of each of CFUs (three per datum point) it was transformed into the logarithm with base 10 (log₁₀ CFUs ml⁻¹). We can see that the higher the concentration of H₂O₂ is the higher the

inhibition effect of growth of the bacterial culture is, so we need to find alternative method that could induce sensitivity of microbes to comparatively low concentrations of H₂O₂.

C. The study of the antibacterial properties of CO₂/H₂O₂ combination on the growth of *E. coli* K-12

The examination of the above mentioned hypothesis shows that the toxic effect of low concentrations of H₂O₂ on *E. coli* K-12 was increased in cell bathing medium enriched with CO₂ for 30 min (maximum) (Fig. 4, (▲)), than in case of non CO₂ enriched bathing medium (Fig. 4, (■)). So fig. 4 shows that the antibacterial effect of H₂O₂ could be elevated with CO₂ enrichment.

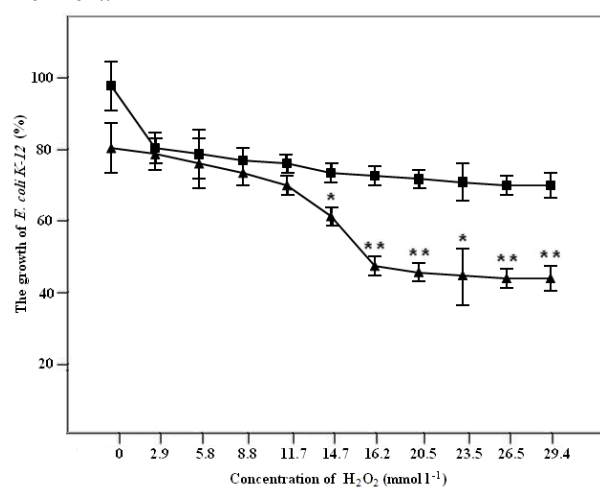


Fig. 4 The effect of different concentrations of H₂O₂ (mmol l⁻¹) on the growth of *E. coli* K-12 (the percent of killed *E. coli* K-12) in 30 min CO₂ enriched bathing medium (▲) and in case of non CO₂ enriched bathing medium (■). p=0.01 (**), p=0.05 (*), n=3.

At the same time it has also been shown that through the antibacterial effect of 5min-CO₂-exposure (minimum) twice could be elevated the effect of 5.2 mmol l⁻¹ H₂O₂ (Fig. 5).

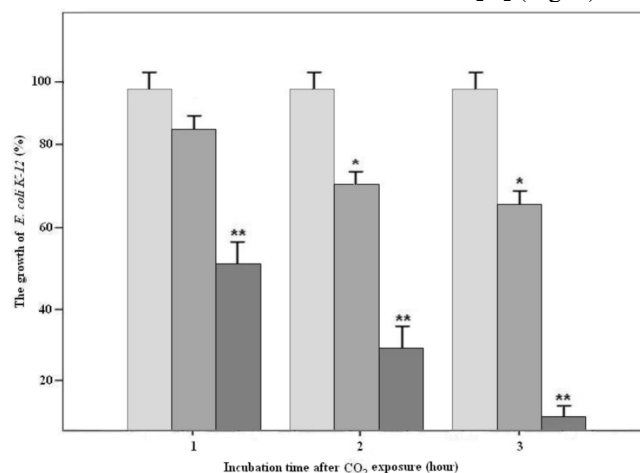


Fig. 5 The ratio of killed *E. coli* K-12 (%) as a function of time-dependent inhibition of the growth of bacterial culture

after 5 min of CO₂ exposure for two times: incubation at 20°C: 5.2 mmol l⁻¹ H₂O₂ containing (1), CO₂ enriched (one time) medium with 5.2 mmol l⁻¹ H₂O₂ (2) and CO₂ enriched (two times) medium with 5.2 mmol l⁻¹ H₂O₂ (3): control or 0 time (■), after 1 h incubation (■) and after 2 h incubation (■). 5 min CO₂ exposure for two times was done before and after 1 h incubation at 20°C and then samples were incubated in thermostat for 2 h with 5.2 mmol l⁻¹ H₂O₂, after that we have done plating and then samples were incubated at 37°C for 22 h. p = 0.01 (**), p = 0.05 (*), n = 3.

So the toxic effect of H₂O₂ is realized by its oxidative properties, consequently, it depends on the initial metabolic activity of microbes [1]. It is suggested that the factor having stimulatory effect on cell metabolism could elevate the toxic effect of H₂O₂ on microbes. On the basis of the fact that CO₂ at comparatively low concentrations [6] has strong stimulatory effect on cell metabolism it could be suggested that cells pretreated by CO₂ could have high sensitivity to toxic effect of H₂O₂ (Fig. 4, 5). It should be mentioned that stimulation of growth occurs because some anabolic reactions involve CO₂ fixation and in the absence of an external source of the gas, CO₂ concentration in cell can be rate limiting for these reactions, with resultant decreased growth rates [15]. The basis of CO₂ inhibition has not been clearly established.

IV. CONCLUSION

The obtained data in the present work clearly show that CO₂ increases the toxic effect of H₂O₂ on microbes and it could be used as a promising tool for water purification in places having higher environmental pollution with CO₂. Although, the obtained data in the present work did not allow us to make final conclusion on the nature of the mechanism through which the CO₂ poisoning metabolism could increase cell membrane sensitivity to H₂O₂, but corresponds to the literature data on CO₂-induced swelling of microbes [4], and our previous data obtained on eukaryotes, on cell swelling-induced increase of a number of chemoreceptors in the membrane [18], allow us to explain CO₂-induced elevation of the toxic effect of H₂O₂ on microbes by the increase of the number of membrane receptors for H₂O₂ in the result of cell swelling. But for final conclusion we need more detailed investigation. So this research is a new approach for decreasing microbial pollution in water, as well as for better understanding of the fundamental mechanisms controlling the microbial growth inhibition depending from environmental factors. We believe that this work will represent the first systematic investigation of the effect of CO₂/H₂O₂ combination at comparatively low concentrations on the reduction of microbial pollution of water.

REFERENCES

[1] A. Hegde, G.K. Bhat, S. Mallya, "Effect of exposure to hydrogen peroxide on the virulence of *Escherichia coli*," *Indian J Med Microbiol*, vol. 26, pp. 8-25, 2008.

[2] A. Mokrini, D. Oussi, S. Esplugas, "Oxidation of aromatic compounds with UV radiation/zone/hydrogen peroxide," *Water Sci. Technol.*, vol. 35, pp. 95-102, 1997.

[3] C.I. Wei, M.O. Balaban, S.Y. Fernando and A. J. Peplow, "Bacterial effect of high pressure CO₂ treatment on foods spiked with *Listeria* or *Salmonella*," *J. Food Prot.*, vol. 54, pp. 189-193, 1991.

[4] E. Debs-Louka, N. Louka, G. Abraham, V. Chabot, K. Allaf, "Effect of Compressed Carbon Dioxide on Microbial Cell Viability," *Appl. and Environm. Microbiolog.*, vol. 65(2), pp. 626-631, 1999.

[5] G. Brandi, M. Fiorani, C. Pierotti, A. Albano, F. Cattabeni, O. Cantoni, "Morphological changes in *Escherichia coli* cells exposed to low or high concentrations of hydrogen peroxide," *Microbiol Immunol.*, vol. 33(12), pp. 991-1000, 1989.

[6] H. Kumagai, C. Hata and K. Nakamura, "CO₂ sorption by microbial cells and sterilization by high pressure CO₂," *Biosci. Biotech. Biochem.*, vol. 61(6), pp. 931-935, 1997.

[7] J.S. Jackson, *Writing the Global Water Crisis Technology and Culture*, vol. 49(3), pp. 773-778, 2008.

[8] J.S. King and L.A. Mabbitt, "Preservation of raw milk by the addition of carbon dioxide," *J. Dairy Res.*, vol. 49, pp. 439-447, 1982.

[9] J.R. Wesley Eckenfelder, "The role of chemical oxidation in waste water treatment process, chemical oxidation technologies for the nineties," *Water Environment & Technology*, pp.1-4, 1992.

[10] J. Zhang, N. Dalal, A. Matthews, "Supercritical carbon dioxide and hydrogen peroxide cause mild changes in spore structures associated with high killing rate of *Bacillus anthracis*," *Journal of Microbiological Methods*, vol. 70(3), pp. 442-451, 2007.

[11] K.A. Presser, D.A. Ratkowsky and T. Ross, "Modeling the growth rate of *Escherichia coli* as a function of pH and lactic acid concentration," *Appl Environ Microbiol*, vol. 63(6), pp. 2355-2360, 1997.

[12] N.M. Dixon and D.B. Kell, "The inhibition by CO₂ of the growth and metabolism of micro-organisms," *J. Appl. Bacteriol.*, vol. 67, pp. 109-136, 1989.

[13] N.R. Asad, L.M. Asad, A.B. Silva, I. Felzenszwalb, A.C. Leitão, "Hydrogen peroxide effects in *Escherichia coli* cells," *Acta Biochim Pol.*, vol. 45(3), pp. 677-90, 1998.

[14] P. Ballestra, A. Abreu Da Silva and J.L. Cuq, "Inactivation of *Escherichia coli* by carbon dioxide under pressure," *J. Food Sci.*, vol. 61, pp. 829-831, 1996.

[15] R. Repaske and M.A. Clayton, "Control of *Escherichia coli* growth by CO₂," *J. Bacteriol.*, vol. 135, pp. 1162-1164, 1978.

[16] R.S. Stepanyan, Zh.R. Alaverdyan, H.G. Oganessian, L.S. Markosyan, S.N. Ayrapetyan, "The Effect of Magnetic Fields on Ion Mutant of *Escherichia coli* K-12 Growth and Division," *Radiational Biology Radioecology*, vol. 40(3), pp. 319-322, 2000.

[17] S. Ayrapetyan, E. Moosavi, V. Martirosyan, "New method for water purification from microbes," AM Patent 20080155, 2008.

[18] S.N. Ayrapetyan and V.L. Arvanov, "On the mechanism of the electrogenic sodium pump dependence on membrane chemosensitivity," *Comp. Biochem. Physiol.*, vol. 64A, pp. 601-604, 1979.

[19] S.N. Ayrapetyan and M.A. Suleymanian, "On the pump induced cell volume changes," *Comp. Biochem. Physiol.*, vol. 64A, pp. 571-575, 1979.

[20] S.N. Ayrapetyan, "On the physiological significance of the pump-induced cell volume changes," *Adv. Physiol. Sci.*, vol. 23, pp. 67-82, 1980.

[21] S.N. Ayrapetyan, R.S. Stepanyan, G.G. Oganessian, A.A. Barsegyan, Zh.R. Alaverdyan, A.G. Arakelyan, L.S. Markosyan, "Effect of Mechanical Vibrations on the Ion Mutant of *Escherichia coli* K-12," *Microbiology*, vol. 70(2), pp. 248-252, 2001.

[22] S.N. Ayrapetyan, "Cell aqua medium as a preliminary target for the effect of electromagnetic fields," *Bioelectromagnetics: Current Concepts*, S. Ayrapetyan and M. Markov, eds., NATO Science Series, Springer Press, the Netherlands, pp. 31-64, 2006.

[23] S. Portier, C. Rochelle, Modeling CO₂ solubility in pure water and NaCl-type waters from 0 to 300°C and from 1 to 300 bar, *Journal of Chemical geology*, 217(3-4), 2005, pp. 187-199.

[24] UNESCO & Bergham Books UNESCO, World Water Assessment Program Water A Shared Responsibility-The United Nations World Water Development Report II, New York, 2006.

[25] Zh.R. Alaverdyan, L.G. Akopyan, L.M. Charyan, S.N. Airapetyan, "Impact of Magnetic Fields on Growth Dynamics and Acid Formation in Lactic Acid Bacteria," *Radiobiology*, vol. 65(2), pp. 241-244, 1996.